

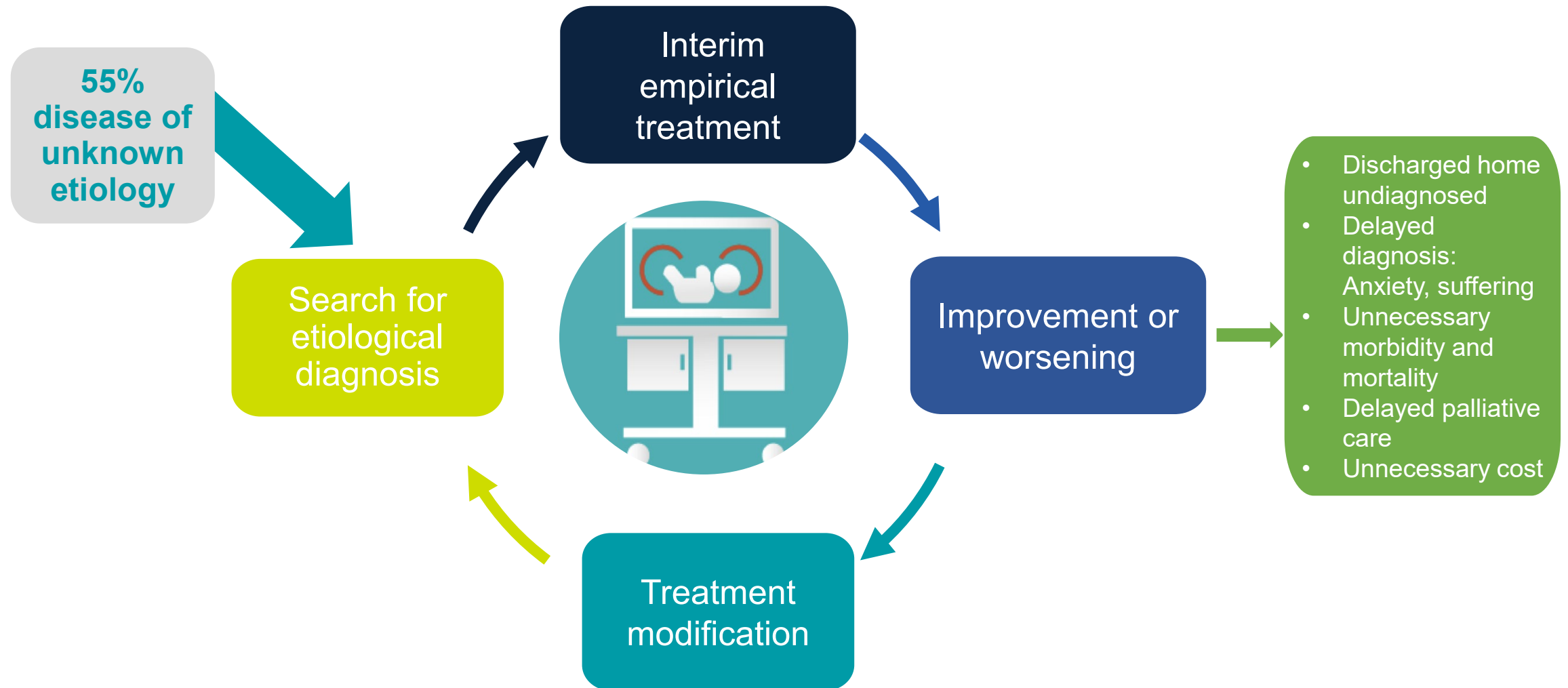
An evaluation of machine intelligence tools to diagnose genetic diseases in critically ill infants

Michelle Clark PhD

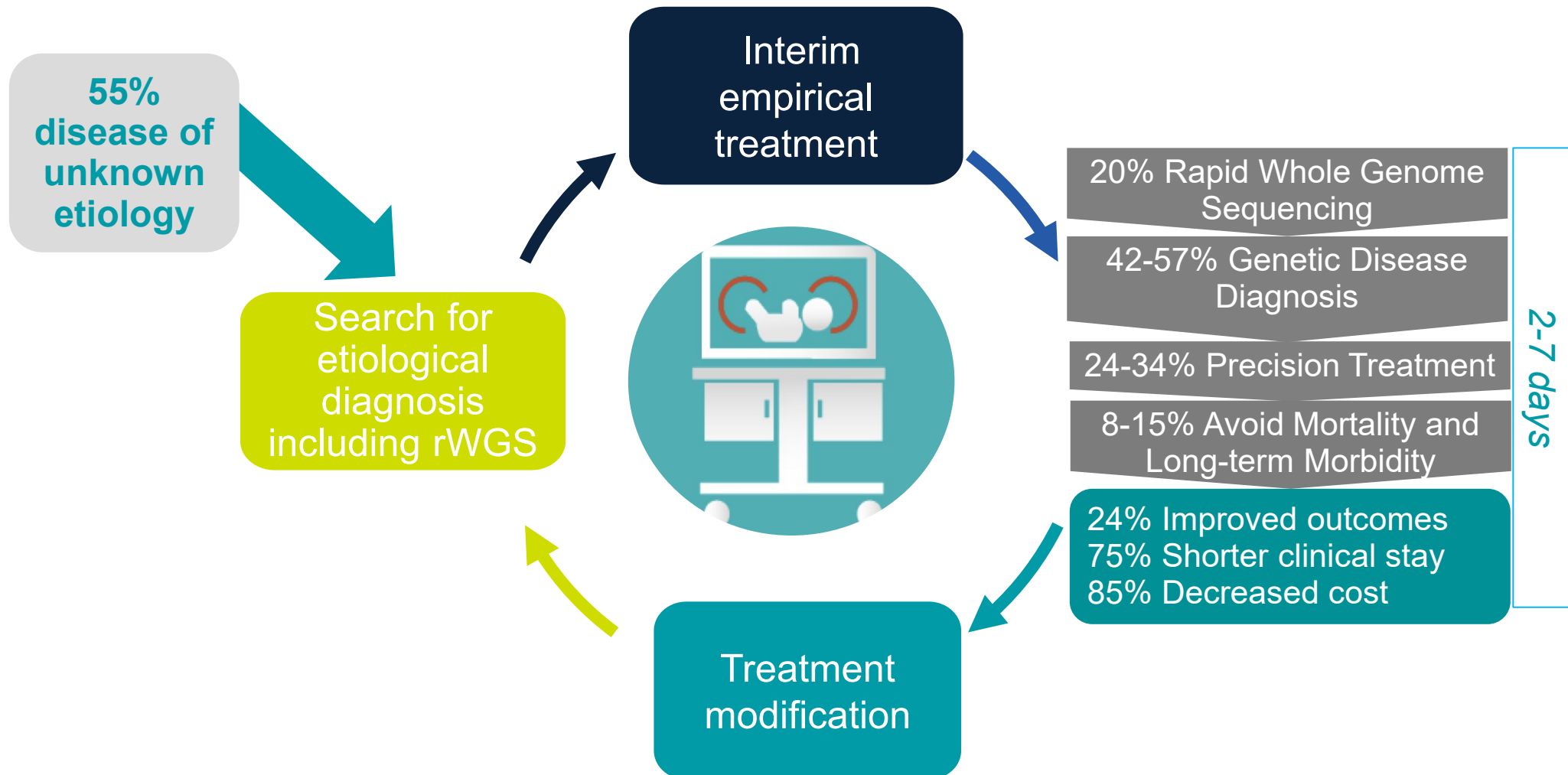
Statistical Scientist, Rady Children's Institute for Genomic Medicine



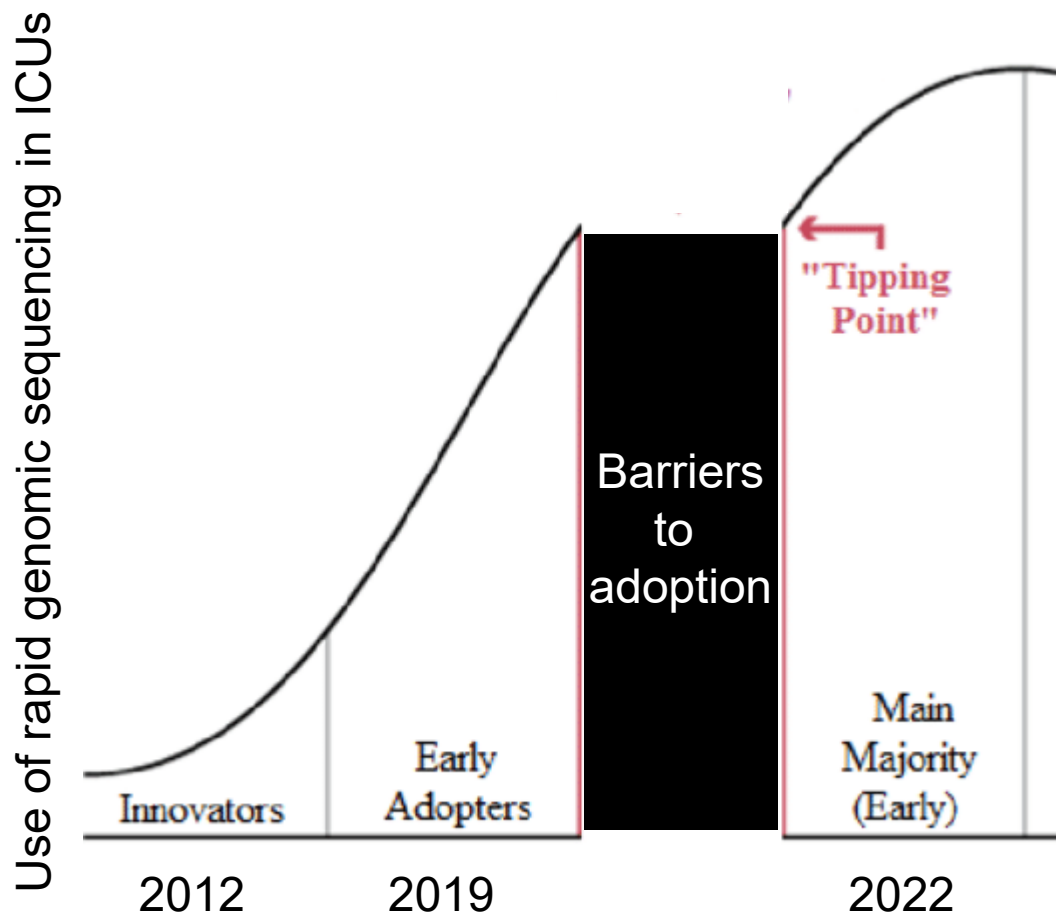
Background



Diagnostic and clinical utility of rapid whole genome sequencing

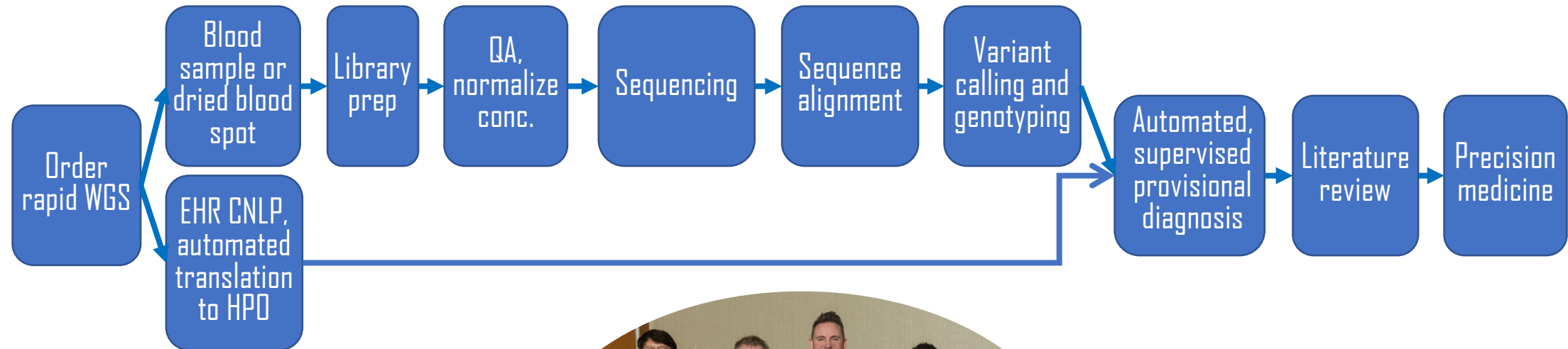


Barriers to broad adoption



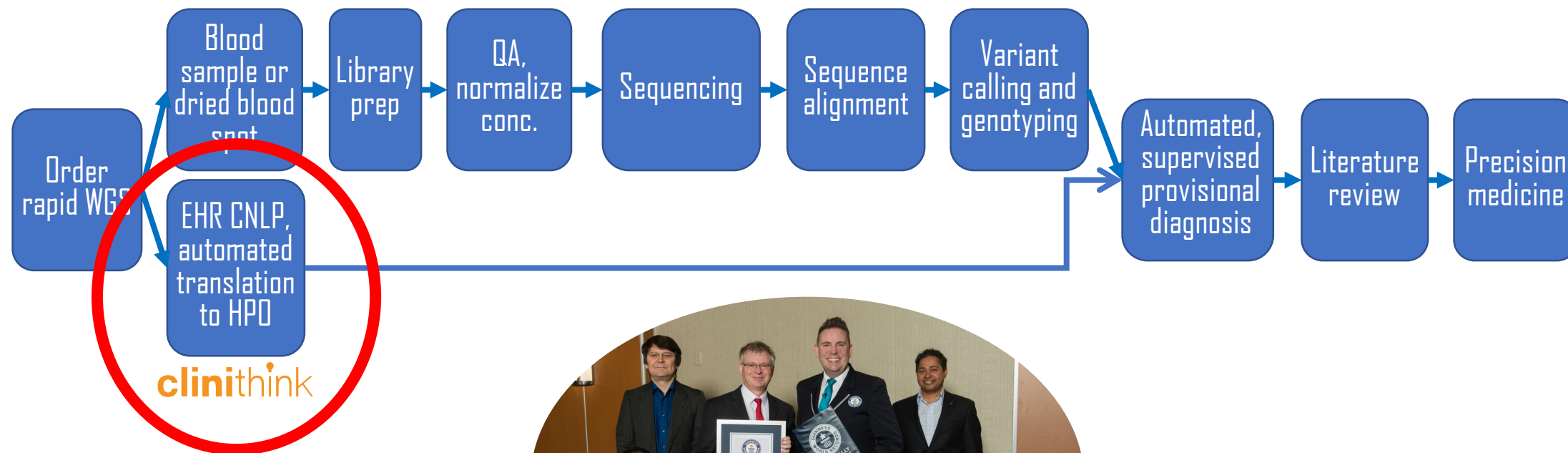
- **Capital & labor intensity of rapid genomic sequencing**
 - Shortage of expert medical geneticists, genetic counselors
 - Not scalable
 - Delays rapid changes in patient care
- **Unfamiliarity with rapid genomic medicine**
 - 13,000 genetic diseases – most of them too rare to have been seen before by pediatricians
- **Insufficient evidence of efficacy**
 - Delayed authorization, failure of reimbursement
- **Many genetic diseases lack effective treatments**
 - Most treatments have not undergone rigorous testing

Solution: automated diagnostic platform using machine intelligence

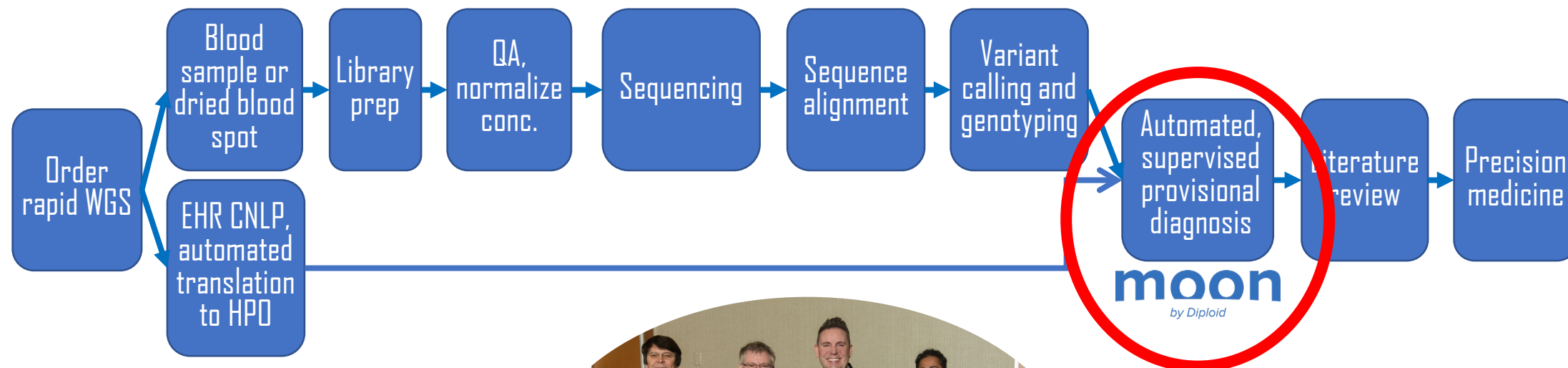


Time from blood draw to provisional diagnosis: 19.5 hours

Automated deep phenotyping



Automated variant interpretation



Evaluation of the automated diagnostic platform

- 1. Retrospective study – 84 children**
- 2. Timed study – 10 children**
- 3. Reanalysis study – 48 children**
- 4. Prospective study – 50 children**

1. Performance in a retrospective cohort:

99% precision, 97% recall

- **95 children with 97 genetic diseases diagnosed manually by rapid whole genome or whole exome sequencing with manually extracted phenotypes and manual interpretation**
- **Excluded incidental findings**
- **99% precision (93 of 94)**
- **97% recall (94 of 97)**

2. Timed study: 100% precision/recall

Mean time savings: 22hrs

Use Type	Retrospective Patients				Prospective Patients													
Subject ID	263	6124	3003	6194	290	352	362	374	7052	412								
Age	8 days	14 years	1 year	5 days	3 days	7 weeks	4 weeks	2 days	17 months	3 days								
Sex	♀	♂	♀	♀	♂	♀	♂	♂	♂	♂								
Abbreviated Presentation	Neonatal seizures	<u>Rhabdo-myolysis</u>	Dystonia, Dev. delay	Hypoglycemia, seizures	Pulmonary hemorrhage, PPHN	Diabetic ketoacidosis	Neonatal seizures	HIE, anemia	Pseudomonal septic shock	Neonatal seizures								
Method	Auto.	Auto.	Auto.	Auto.	Auto.	Std.	Auto.	Std.	Auto.	Std.	Auto.	Std.	Auto.	Std.	Auto.	Std.	Auto.	Std.
Number of Phenotypic Features	51	115	148	14	2	257	4	103	4	65	1	112	6	124	3	33	1	
Molecular Diagnosis	Early Infantile Epileptic Encephalopathy 7	Glycogen Storage Disease V	<u>Dopa-Responsive Dystonia</u>	None	None	Permanent neonatal diabetes mellitus	None	None	X-linked <u>agammaglobulinemia 1</u>	Benign familial neonatal seizures 1								
Gene and Causative Variant(s)	<i>KCNQ2</i> c.727C>G	<i>PYGM</i> c.2262delA c.1726C>T	<i>TH</i> c.785C>G c.541C>T	None	None	<i>INS</i> c.26C>G	None	None	<i>BTK</i> c.974+2T>C	<i>KCNQ2</i> c.1051C>G								
Sample/Library Prep (hours)	3:20	2:55	2:24	2:22	2:10	23:54	2:12	22:05	2:13	15:42	2:31	18:30	3:30	10:10	4:30	12:10	3:05	23:50
NovaSeq Loading (hours)	0:20	0:17	0:16	0:20	1:38	0:20	0:29	0:22	0:30	0:53	0:15	2:30	0:45	0:35	1:00	1:00	0:20	0:53
2x101 nt Sequencing (hours)	15:36	15:31	15:34	15:27	15:26	24:13	15:25	24:08	15:21	22:44	15:17	33:36	15:17	21:07	15:19	22:46	15:58	21:00
1 ^o & 2 ^o Analysis (hours)	1:03	1:02	0:59	0:59	1:07	3:05	1:00	1:57	1:01	2:30	1:02	2:30	1:02	2:30	1:09	2:25	1:24	2:24
3 ^o Analysis Processing (hours)	0:06	0:05	0:07	0:05	0:06	0:15	0:08	0:14	0:06	0:15	0:05	0:15	10:28	0:16	0:06	0:16	0:06	0:16
Total (hours)	20:25	19:56	19:20	19:14	20:42	56:03	19:29	48:46	19:11	42:04	19:10	57:21	31:02	34:38	22:04	38:37	20:53	48:23

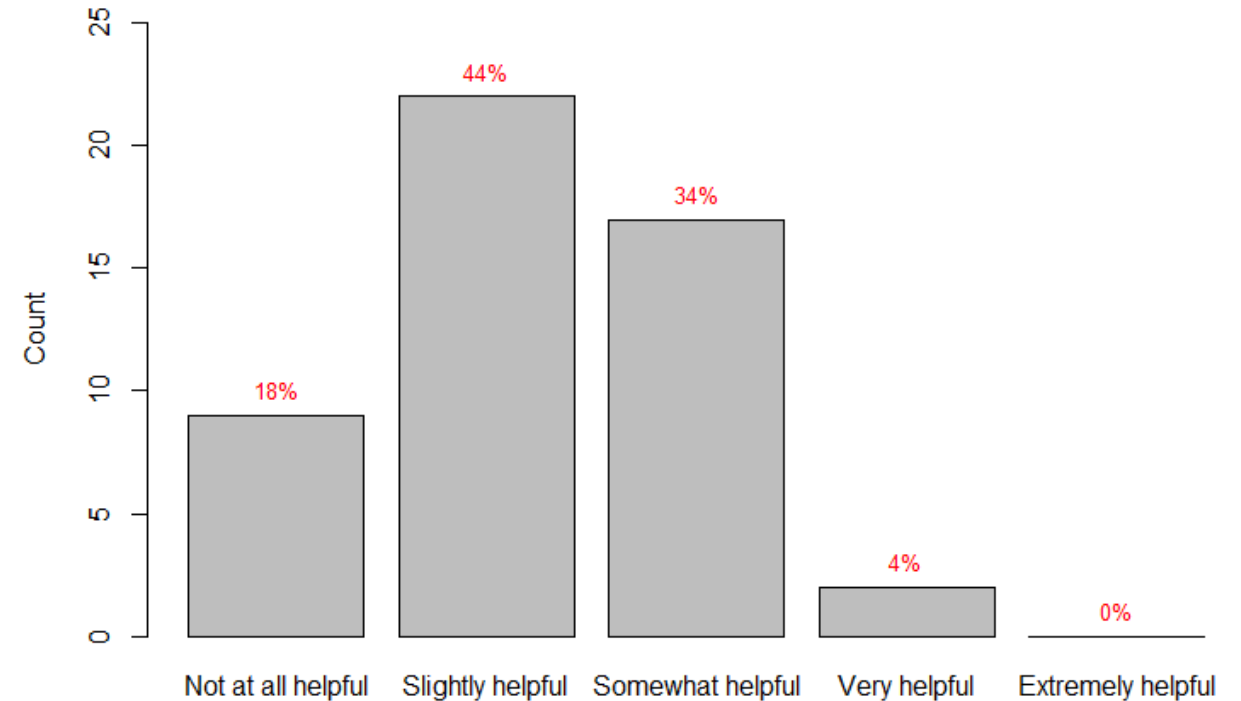
3. Reanalysis study: 4.2% diagnostic yield

- Automation of these reanalysis steps reduced the number of variants under consideration by an average of 99.9%.
- In two cases, diagnoses were made upon reanalysis, representing a yield of 4.2% (2 of 48).
- Four additional cases were flagged with a possible diagnosis to be considered during periodic reanalysis.
- An untrained analyst identified these six diagnoses with specificity = 0.83 and sensitivity = 0.76.

4. Prospective performance: 100% recall

- Out of 50 patients, the standard diagnostic workflow resulted in 16 (32%) diagnoses
- Automated analysis correctly diagnosed all 16 patients (100% recall)
- In addition to the standard workflow, analysts found automation to be very helpful in 4% of cases




“How helpful was automated analysis in addition to the standard workflow?”



Rady Children's Institute for Genomic Medicine – the clinical lab perspective

- **Hesitation when machine intelligence tools undergo rapid updates**
- **Goes against how clinical lab directors were trained to validate tools**
- **Need sufficient warning prior to updates**
- **Request increased transparency**

Moon's response to requests for transparency

BTD ▾ 3:15,677,022 ▾ rs397514336 ClinVar	GT ref: G	stop gained p.Glu48* 3 publications	Biotinidase deficiency AR - compound heterozygous	  
HBB ▾ 11:5,248,232 ▾ rs334 ClinVar	AA ref: T	missense p.Glu7Val 272 publications	Sickle cell anemia AR	  
BTD ▾ 3:15,686,693 ▾ rs13078881 ClinVar	GC ref: G	missense p.Asp446His 78 publications	Biotinidase deficiency AR - compound heterozygous	  

Effect	Transcript	Effect	pnotation	cnotation	Exon rank
	ENST332509	missense	p.Arg591Trp	c.1771C>T	13/17
	ENST335539	missense	p.Arg537Trp	c.1609C>T	12/16
	ENST402064	missense	p.Arg537Trp	c.1609C>T	12/16

Protein prediction	
--------------------	--

Gene region	
-------------	--

Frequency	0.0032%	0	1
	GNOMAD	HOMOZYGOTES	HETEROZYGOTES
	0.0407%		
	DIPLOID		

Quality	77	37,40	99
	DEPTH	ALLELE DEPTH	GENOTYPE QUALITY

Reported variants

Position	Genotype	Gene	Disorder
22:38511635	G/A	PLA2G6	Infantile neuroaxonal dystrophy 1
22:38512190	G/A	PLA2G6	Infantile neuroaxonal dystrophy 1

Variant discussion

PLA2G6 ▾

22:38,511,635 ▾

GA

ref: G

stop gained

p.Arg645*

Infantile neuroaxonal dystrophy 1

AR

Note

Two variants, a novel stop gained variant and a novel stop gained variant, were detected in heterozygous state in the *PLA2G6* gene (ENST332509: c.1933C>T; p.Arg645* and 332509: c.1933C>T; p.Arg645*). Parental DNA analysis is required to establish a compound heterozygous state of these two variants.

Mutations in *PLA2G6* have been shown to cause Infantile neuroaxonal dystrophy 1 (MIM: 256600), an autosomal recessive condition. The reported clinical phenotype of this patient overlaps with the manifestations of this condition regarding neurodegeneration, developmental regression, nystagmus, spastic tetraplegia, and cerebellar atrophy. The typical age of onset of Infantile neuroaxonal dystrophy 1 ranges from 0 to 10, which is in line with the reported age of onset in this patient (1 y.). Further clinical evaluation of the patient will give more insight into the phenotypic overlap with Infantile neuroaxonal dystrophy 1.

The detected variant causes stop gained. It is absent from gnomAD and absent from dbSNP, but has not previously been associated with disease. Parental DNA analysis (trio analysis) and DNA analysis of other (un)affected relatives, could establish co-segregation of this variant with the reported clinical phenotype.

Classification

Unassigned ▾

References

Enter PubMed ID or PubMed URL

Add publication

The clinical lab perspective continued

- **High sensitivity with automation, but unsure about sensitivity**
 - **Trust will come from large studies from other groups of hundreds of thousands of cases**
- **Development of publically available benchmarks to validate methods after every update**
 - **Example: Genome in a Bottle for clinical validation of genome sequencing**

Conclusion

- **Although the automated diagnostic system is “hands-free”, it’s supervised at every step by expert bioinformaticians, clinical medical geneticists and clinical lab directors.**
- **May enable effective first-tier, provisional diagnoses or automated re-analysis of unsolved cases**
- **Wide-spread adoption would allow valuable cognitive resources of molecular laboratory directors and analysts to be reserved for difficult cases, manual curation of variants, and clinical report generation**

Acknowledgments

Michelle M. Clark¹, Kiely N. James¹, Amber Hildreth^{1,2,3}, Sergey Batalov¹, Yan Ding¹, Shimul Chowdhury¹, Kelly Watkins¹, Katarzyna Ellsworth¹, Brandon Camp¹, Cyrielle I. Kint⁴, Calum Yacoubian⁵, Lauge Farnaes^{1,2}, Matthew N. Bainbridge^{1,6}, Curtis Beebe⁷, Joshua J. A. Braun¹, Margaret Bray⁸, Jeanne Carroll^{1,2}, Julie A. Cakici¹, Sara A. Caylor¹, Christina Clarke¹, Mitchell P. Creed⁹, Jennifer Friedman^{1,10}, Alison Frith⁵, Richard Gain⁵, Mary Gaughran¹, Shauna George⁷, Sheldon Gilmer⁷, Joseph Gleeson^{1,10}, Jeremy Gore¹¹, Haiying Grunenwald¹², Raymond L. Hovey¹, Marie L. Janes¹, Kejia Lin⁷, Paul D. McDonagh⁸, Kyle McBride⁷, Patrick Mulrooney¹, Shareef Nahas¹, Daeheon Oh¹, Albert Oriol⁷, Laura Puckett¹, Zia Rady¹, Martin G. Reese¹³, Julie Ryu^{1,2}, Lisa Salz¹, Erica Sanford^{1,2}, Lawrence Stewart⁷, Nathaly Sweeney^{1,2}, Mari Tokita¹, Luca Van Der Kraan¹, Sarah White¹, Kristen Wigby^{1,2}, Brett Williams⁵, Terence Wong¹, Meredith S. Wright¹, Catherine Yamada¹, Peter Schols⁴, John Reynders⁸, Kevin Hall¹², David Dimmock¹, Narayanan Veeraraghavan¹, Thomas Defay⁸, Stephen F. Kingsmore^{1*}.

¹Rady Children's Institute for Genomic Medicine, San Diego, CA 92123, USA;

²Department of Pediatrics, University of California San Diego, San Diego, CA 92093, USA;

³Department of Pediatrics, University of Washington, Seattle, WA 98195, USA;

⁴Diploid, 3001 Leuven, Belgium;

⁵Clinithink Ltd., London N1 6DR, UK;

⁶Codified Genomics, LLC, Houston, TX 77033, USA;

⁷Rady Children's Hospital, San Diego, CA 92123, USA;

⁸Alexion Pharmaceuticals, Inc., New Haven, CT 06510, USA;

⁹University of Kansas School of Medicine, Kansas City, MO 66160, USA;

¹⁰Department of Neurosciences, University of California San Diego, San Diego, CA 92093, USA;

¹¹Tessella, Needham, MA 02494, USA;

¹²Illumina, Inc., San Diego, CA 92122, USA;

¹³Fabric Genomics, Inc., Oakland, CA 94612, USA.

Support:
NICHD
NHGRI
Illumina
Alexion
Diploid
Clinithink
Fabric